

Future treatment strategies in phenylketonuria [☆]

Francjan J. van Spronsen ^{a,*}, Gregory M. Enns ^b

^a Section of Metabolic Diseases, Beatrix Children's Hospital, University Medical Centre of Groningen, PO Box 30.001, 9700 RB, Groningen, The Netherlands

^b Division of Medical Genetics, Department of Pediatrics, Lucile Packard Children's Hospital, Stanford University, 300 Pasteur Drive, H-315, Stanford, CA 94305-5208, USA

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ABSTRACT

Phenylketonuria (PKU) was the first inherited metabolic disease in which treatment was found to prevent clinical features of the disorder; dietary management was established almost 60 years ago. The institution of a low-phenylalanine (Phe) diet in the first few weeks of life was made possible by Guthrie neonatal screening, which further increased effectiveness of therapy. Indeed, neonatal diagnosis of PKU followed by institution of a low-Phe diet has been a remarkable success in preventing the devastating brain damage associated with untreated PKU. Nevertheless, significant difficulties exist in caring for PKU patients, including problems with adhering to the prescribed dietary regimen and the presence of neurocognitive deficits despite therapy. During the past few years, several ideas for new treatment strategies have emerged. This review aims to address these treatment strategies based on theoretical considerations of the biochemistry and pathogenesis of PKU. Recent times have seen the introduction of a wide array of novel treatments currently in clinical use, including more palatable medical foods, glycomacropeptide, large neutral amino acids, and tetrahydrobiopterin. Human trials are underway using an enzymatic therapeutic approach, while preclinical work continues in the fields of gene and cellular therapy. These therapeutic strategies propose to treat PKU at various levels, including nutritional intake, gut, liver, and blood–brain barrier, and have the potential to further improve outcome in PKU.

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Introduction

Phenylketonuria (McKusick 261600, PKU) is one of the best-known examples of inherited metabolic diseases. It was the first disorder in which a toxic agent, phenylalanine (Phe), was identified to cause severe mental retardation. It was also the first known defect in amino acid metabolism that involved the discovery of Phe-to-tyrosine metabolism and the primary enzyme deficiency of hepatic phenylalanine hydroxylase (PAH; EC1.14.16.1). More

Abbreviations: PKU, phenylketonuria; Phe, phenylalanine; PAH, phenylalanine hydroxylase; BBB, blood–brain barrier; IQ, intelligence quotient; GMP, glycomacropeptide; PAL, phenylalanine-ammonia lyase; LNAA, large neutral amino acids; BH₄, tetrahydrobiopterin; FDA, Food and Drug Administration; EMEA, European Medicines Agency; rAAV, recombinant adeno-associated virus.

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* Corresponding author.

E-mail addresses: f.j.van.spronsen@bkk.umcg.nl (F.J. van Spronsen), greg.enns@stanford.edu (G.M. Enns).

importantly, PKU was the first inherited metabolic disease in which treatment was found to prevent clinical features of the disorder. The dietary management of PKU was established almost 60 years ago [1], with the first effects of treatment published in 1953 [2]. Many reviews have emerged since then, with one of the most recent reviews addressing the sizeable collection of knowledge accumulated to date [3].

During the past few years, several ideas for new treatment strategies have emerged. This paper aims to address these treatment strategies based on theoretical considerations of the biochemistry and pathogenesis of PKU. These therapeutic strategies aim to treat PKU at various levels, including nutritional intake, gut, liver, blood–brain barrier (BBB), and other areas in the body (i.e., gene therapy) (Fig. 1).

The successes and challenges of dietary treatment

High levels of Phe are considered toxic, and since Phe is not synthesized by the body, the low-Phe diet was introduced. The effectiveness of treatment was further increased by starting the low-Phe diet in the first few weeks of life. This was made possible by the introduction of Guthrie neonatal screening, which screened for high Phe concentrations in blood taken by heel puncture [4]. In principle, the diet restricts the amount of Phe intake to the amount needed for protein anabolism. In other words, the amount

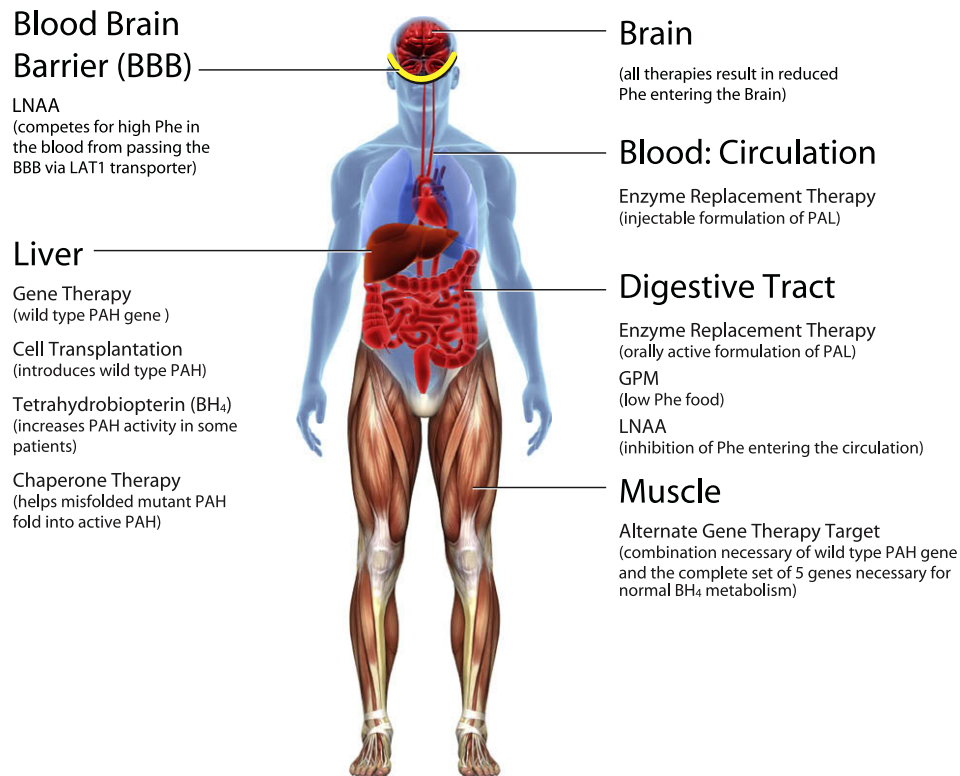


Fig. 1. The figure presents the different treatment options available in practice or in theory, looking at different locations in the body.

of Phe necessary for protein synthesis is provided by normal nutritional protein intake. The amount of Phe normally converted into tyrosine, which can represent up to 90% of Phe intake, is replaced by medical food substitutes. This treatment has proven to result in grossly normal cognition. However, when compared to siblings or healthy age-related control groups, there is always a small but consistent gap in intelligence quotient (IQ) scores [5–7]. In addition, neuropsychometric testing has identified problems particularly in executive functioning at various ages [8–13], although the hypothesized central role that prefrontal lobe damage plays in such findings has been questioned [14]. There is also a risk of regression of neurocognitive function if patients are not treated in adulthood, possibly related to brain white matter abnormalities or impaired synthesis of brain catecholamines [15–17]. Therefore, there is increasing consensus for the need of lifelong dietary treatment. Despite such consensus, dietary management practices are notoriously variable throughout the world. Although Phe restriction is the mainstay of PKU treatment worldwide, significant differences exist in recommended blood Phe levels in both children and adults [18–23].

Apart from the fact that the diet does not result in a completely normal outcome, the diet is not easy to comply with, resulting in a social burden, which may result in various social problems [24]. Additionally, treatment with special medical foods is associated with risks of nutritional deficiencies, especially concerning vitamin B₁₂, vitamin D, calcium, iron, and unsaturated long chain fatty acids [25–27]. Such deficiencies may result in neurological problems and bone density-related issues [28,29]. Therefore, we need to consider additional therapeutic possibilities in the treatment of PKU patients.

At the level of dietary intake

During the past few years, many issues concerning protein substitutes used to treat PKU have been improved, including content,

palatability, and practical use of the products. Apart from this, there is now some experience with a protein that is almost free of Phe. This protein, glycomacropeptide (GMP), is derived from goat milk during cheese making and has a better taste than protein substitutes. A drawback of this treatment is that it cannot completely replace the need for supplemental protein substitute because it is not completely free of Phe. In addition, some amino acids must be added because natural GMP does not include other amino acids such as tyrosine and tryptophan [30]. Still, it is a very interesting and palatable product that is a good addition to the PKU treatment arsenal, especially given how difficult it is for many patients to adhere to the low-Phe diet [31,32].

Another advancement that has the potential to improve dietary adherence is the development of a portable Phe monitoring device [33,34]. PKU patients often have to wait days to weeks to obtain the results of a blood Phe test. The ability to monitor blood Phe levels in real time could serve as an important tool to help motivate patients to follow dietary recommendations more closely.

At the gut level

Although the gut does not contain the essential pathways for Phe metabolism, it may help to treat PKU in two different ways. First, the enzyme phenylalanine-ammonia lyase (PAL, EC 4.3.1.5) is active at this site if taken orally in a non-absorbable and protected form [35–37]. To increase the activity and mask PAL from the host immune system, the injectable form of PAL is PEGylated. PEGylation is the process of attachment of polyethylene glycol polymer chains to PAL [38,39]. In its present PEGylated formulation the maximal effect is likely in the vascular space. PAL is an enzyme that metabolizes Phe into the non-toxic compound trans-cinnamic acid and ammonia. The small amount of ammonia released is easily converted into urea. Preclinical studies in murine models of PKU using gavage feeding or intraperitoneal injection of PAL or subcutaneous injection of a PEGylated enzyme preparation (PEG-PAL conjugates)

have been promising, showing lowered Phe levels in both brain tissue and vascular space and reduced manifestations of disease [36,38]. In addition, PEGylation allows for subcutaneous depot injection of PEG-PAL, a form of enzyme delivery that has the potential to be relatively long-acting. Even with such enzyme therapy tyrosine remains an essential amino acid; the amount of required supplementation of this amino acid during PAL treatment is a matter for further study. The first clinical trial of PEG-PAL was started in the USA in 2008 (PAL-001, NCT00634660).

Second, supplementation with large neutral amino acids (LNAA) decreases the blood Phe concentration [40]. This is a secondary effect of LNAA, while the primary effect is to decrease the brain Phe concentration. The concept behind LNAA treatment is that Phe and other LNAA share the same transport system, creating competitive inhibition of the transport of LNAA with each other [40,41]. Therefore, LNAA supplementation counteracts the effects of high Phe exposure to the transport system that delivers LNAA across membranes from gut to blood or from blood to brain [42]. In the gut, Phe and other LNAA compete for transport from the apical to the basolateral membrane [43]. When PKU patients are off dietary therapy, the blood Phe concentration is very high. In such instances, large amounts of LNAA may reduce the influx of Phe to the brain, as was clearly shown by the study of Pietz and coworkers [41]. However, at the gut level, the Phe concentration is more or less normal, so that supplying large amounts of LNAA may not decrease Phe transport to the blood appreciably, although a recent study has shown a significant drop in blood Phe (39%) following treatment of PKU patients with LNAA for one week [44]. Therefore, the effect of LNAA supplementation on blood Phe concentration might be a result of other factors, including stimulating anabolism or potentially improving the competitive effect of LNAA resulting from decreased natural protein intake, rather than directly influencing transport mechanisms [45]. This is in line with the finding that the blood Phe concentration decreases when amino acid supplements are given more frequently in conjunction with non-LNAA [46,47].

In the liver

The essential part of Phe metabolism takes place in the liver. This was conclusively shown to be true after a boy with PKU and unrelated end-stage liver disease underwent liver transplantation; complete correction of the metabolic phenotype followed [48]. Normally, the entire set of enzymes is active in hepatocytes, including PAH as well as the five enzymes involved in the synthesis and recycling of (6R)-L-erythro-5,6,7,8-tetrahydrobiopterin (BH₄). In the first years of treatment, although dietary therapy was successful in preserving intelligence in the vast majority of PKU patients, some were found to respond poorly to dietary intervention despite adequate control of their blood Phe levels. These patients were later shown to have BH₄ deficiency due to a defect in either the synthesis or recycling of BH₄ [49,50]. A BH₄ loading test was developed to distinguish the patients with a defect in BH₄ metabolism from patients with deficient PAH activity [51]. The patients with a defect in BH₄ metabolism could also be detected by analysing urine pterins and measuring the activity of one of the enzymes involved with the recycling of BH₄ [50,52]. In 1999, however, two things changed: the effectiveness of BH₄ increased by a change in the formulation, while the dose was also increased from 10 to 20 mg/kg body weight. This resulted in detecting a third group of patients, the so-called BH₄-responsive patients. These patients neither have a defect in BH₄ metabolism nor BH₄ deficiency, but do exhibit an increase of *in vivo* PAH capacity with large doses of BH₄.

Since then, some patients with a deficiency of the PAH enzyme were found to be responsive to BH₄ [53–55]. These patients usually

have a milder form of PAH deficiency. While a large number of patients with a mild degree of PAH deficiency are responsive to BH₄, a smaller number of patients with more severe PAH deficiency also clearly show responsiveness [56,57]. The exact proportion of BH₄-responders who have a more severe phenotype is not yet known, due to selection bias associated with initial clinical trials [58,59]. However, even some patients with a mutation in the gene encoding PAH, resulting in a clear lack of enzyme activity, might show some response [60]. Therefore, while it is of importance to make clear to families that many patients will not be responsive, every classic PKU patient should be offered testing [57]. Experience shows that most of the patients responsive to BH₄ still need at least some restriction of natural protein and continued use of low-Phe medical foods. Following the initial demonstration of BH₄ efficacy in mild PKU patients in 1999 [53], 6R-BH₄ (sapropterin dihydrochloride, Kuvan®) has been developed from a chemical compound into an FDA and EMEA approved drug (December 2007 and 2008, respectively). The product has been launched in the US and in a growing number of European countries.

At the level of genes and cells

After the detection of the PAH gene by Kwok et al. [61] and the studies of Ledley et al. [62] showing that only about 10% enzyme activity is necessary for normal Phe metabolism in mice, further experimental protocols emerged to treat PKU with gene therapy [61–63]. However, after the unfortunate death of a patient with another inborn error of metabolism (ornithine transcarbamylase deficiency) [64], it became clear that there were important issues to be addressed before a gene therapy strategy could be used widely in PKU patients. Eighteen hours following gene transfer, the subject developed a systemic inflammatory response syndrome with multiple organ system failure, ultimately leading to death [64]. Nevertheless, more recent studies in murine models of PKU are promising. Long-term correction of murine hyperphenylalaninemia associated with no adverse effects has been achieved by the administration of a recombinant adeno-associated virus (rAAV) vector [65]. AAV vectors appear to be less prone to cause an immune reaction that would result in rejection of transfected hepatocytes when compared to the initial vectors used in gene therapy experiments [66]. Such advances in viral vector design, especially related to improvements in AAV technology, have led to human gene therapy trials for other inborn errors of metabolism, including Canavan disease, late-infantile neuronal ceroid lipofuscinosis, α_1 -antitrypsin deficiency, and lipoprotein lipase deficiency [67].

Hepatocyte transplantation in PKU is also under investigation, but, in order to be effective, donor cells need to have a selective growth advantage over native hepatocytes [66]. Hepatocyte transplantation has been attempted in a number of preclinical studies using various animal models of inborn errors of metabolism, as well as in humans with metabolic disorders such as urea cycle defects, glycogen storage disorders, α_1 -antitrypsin deficiency, and Refsum disease. This cellular approach has not yet been attempted in PKU. Cell-based therapies using stem cells or more differentiated progenitor cells may represent the future of cell transplantation for treatment of metabolic liver disease, and eventually may be a viable option for treating PKU patients [68].

In the muscle

Apart from the reports on liver transfection, there are some innovative studies on muscle as a target for gene therapy [69]. In order to introduce the Phe hydroxylating system into tissue other than the liver, gene delivery must include not only the PAH enzyme but also transgenes that encode the complete enzyme

system necessary to synthesise and recycle BH₄. Despite such a daunting technical challenge, Ding et al. [69] have shown this to be possible in mice. One tremendous advantage of muscle gene therapy is the ease of administration of the therapeutic vectors when compared to liver-directed gene therapy.

At the brain level

Notwithstanding the good results of treatment and the clear relationship between blood Phe concentrations and cerebral outcome, the pathogenesis of brain damage in PKU is still largely not understood [70,71]. The fact that the deleterious effects of PKU are almost completely restricted to the brain, in addition to the fact that the brain, unlike other tissues, is separated from the blood by the BBB, lends support to the hypothesis that the BBB is of central importance to disease pathogenesis.

The studies of Möller et al. [72], Weglage et al. [73], Koch [74], and Moats et al. [75] showed that some patients with untreated classical PKU who consequently have very high blood Phe concentrations do not show evidence of significantly impaired cognitive function. Despite high blood Phe concentrations, such individuals showed almost normal cerebral Phe concentrations [73–75]. These findings again point to the possible importance of the BBB in the pathogenesis of brain disease in PKU. In particular, polymorphisms in the gene encoding the LNAA transport system (the neutral amino acid transporter 1, LAT1) might play a major role in determining susceptibility to brain injury in PKU patients. For the LNAA transport across the BBB, alterations in the 4F2hc/LAT1 complex have been suggested to be a possible explanation for the phenomenon found by Weglage et al. [72,73]. However, no sequence variants that would be expected to result in a structural change of this transporter were detected in 13 PKU patients who had demonstrated a low ratio of brain to blood Phe concentrations [72], so the cause of the discordance between blood and brain Phe levels in some patients is still unknown.

As already mentioned above, LNAA supplementation is an approach that may lower blood Phe levels in some instances, although the major effect appears to be on brain Phe levels [76]. Christensen [77] proposed in 1953 that high blood Phe concentrations could interfere with transport of other LNAA into the brain. In 1976, Andersen et al. [78] demonstrated the potential for LNAA to lower brain Phe levels in a rat model. Although other studies also concentrated on altering brain amino acid and catecholamine composition in PKU through a modified dietary approach [79–81], it was not until the groundbreaking work of Pietz et al. [41] that LNAA supplementation was studied in earnest in PKU patients. Provision of large amounts of LNAA in the diet resulted in clearly decreased brain Phe concentrations and improved neurophysiological function [41]. A recent prospective, double-blind, crossover study using LNAA supplementation showed no correlation between plasma and brain Phe levels. However, LNAA supplementation improved executive functioning, especially in the domains of verbal generativity and cognitive flexibility. The authors concluded that use of LNAA in patients who are compliant with diet therapy is of limited value, but LNAA supplementation may benefit those who are unable to comply with the dietary restrictions necessary to treat PKU [82].

The list of known amino acid transporters continues to grow, so new studies are necessary to identify genes and polymorphisms that have the potential to affect brain Phe levels. Once novel transporter genes are found, our understanding of the phenotypic variability in patients, including which patients may need either less strict treatment or no treatment at all, should increase.

Finally, BH₄ appears to have the potential to cross the BBB, at least at high doses, and hence might directly affect neurotransmit-

ter levels. Peripherally administered BH₄ was found to enter the brain in rats and monkeys [83,84]. When BH₄ or 6-methyltetrahydrobiopterin was given orally along with ascorbate or intravenously to patients with defective BH₄ synthetic pathways at doses of 10–20 mg/kg, substantial amounts were detected in the cerebrospinal fluid [85]. On the other hand, a recent study of murine tyrosine hydroxylase activity during BH₄ supplementation showed that only high doses (100 mg/kg) resulted in a significant increase of brain tyrosine hydroxylase activity [86]. Therefore, Thöny et al. [86] considered it unlikely that current therapeutic regimens using BH₄ to treat PKU in humans (with doses typically <20 mg/kg) would have much of an effect on brain neurotransmitters.

Conclusion

Neonatal diagnosis of PKU followed by institution of a low-Phe diet has been a remarkable success in preventing the devastating brain damage associated with untreated PKU. Nevertheless, significant difficulties exist in caring for PKU patients, including problems with adhering to the prescribed dietary regimen and the presence of neurocognitive deficits despite therapy. Recent times have seen the introduction of a wide array of novel treatments currently in clinical use, including more palatable medical foods, GMP, LNAA, and BH₄. Human trials are underway using an enzymatic therapeutic approach (PEG-PAL), while preclinical work continues in the fields of gene and cellular therapy. Taken together, the future of PKU treatment has never looked brighter.

References

- [1] L.I. Woolf, Excretion of conjugated phenylacetic acid in phenylketonuria, *Biochem. J.* 49 (1951) ix–x.
- [2] H. Bickel, J. Gerrard, E.M. Hickmans, Influence of phenylalanine intake on phenylketonuria, *Lancet* 265 (1953) 812–813.
- [3] C.N. Sarkissian, A. Gamez, C.R. Scriver, What we know that could influence future treatment of phenylketonuria, *J. Inherit. Metab. Dis.* 32 (2009) 3–9.
- [4] R. Guthrie, A. Susi, A simple phenylalanine method for detecting phenylketonuria in large populations of newborn infants, *Pediatrics* 32 (1963) 338–343.
- [5] I. Smith, O.H. Wolff, Natural history of phenylketonuria and influence of early treatment, *Lancet* 2 (1974) 540–544.
- [6] R. Koch, C. Azen, E.G. Friedman, M.L. Williamson, Paired comparisons between early treated PKU children and their matched sibling controls on intelligence and school achievement test results at eight years of age, *J. Inherit. Metab. Dis.* 7 (1984) 86–90.
- [7] K. Fishler, C.G. Azen, E.G. Friedman, R. Koch, School achievement in treated PKU children, *J. Ment. Defic. Res.* 33 (1989) 493–498.
- [8] M.C. Welsh, B.F. Pennington, S. Ozonoff, B. Rouse, E.R. McCabe, Neuropsychology of early-treated phenylketonuria: specific executive function deficits, *Child Dev.* 61 (1990) 1697–1713.
- [9] P. Griffiths, N. Ward, A. Harvie, F. Cockburn, Neuropsychological outcome of experimental manipulation of phenylalanine intake in treated phenylketonuria, *J. Inherit. Metab. Dis.* 21 (1998) 29–38.
- [10] S. Huijbregts, L. de Sonnevile, R. Licht, J. Sergeant, F. van Spronsen, Inhibition of prepotent responding and attentional flexibility in treated phenylketonuria, *Dev. Neuropsychol.* 22 (2002) 481–499.
- [11] S.C. Huijbregts, L.M. de Sonnevile, R. Licht, F.J. van Spronsen, P.H. Verkerk, J.A. Sergeant, Sustained attention and inhibition of cognitive interference in treated phenylketonuria: associations with concurrent and lifetime phenylalanine concentrations, *Neuropsychologia* 40 (2002) 7–15.
- [12] S.C. Huijbregts, L.M. de Sonnevile, F.J. van Spronsen, R. Licht, J.A. Sergeant, The neuropsychological profile of early and continuously treated phenylketonuria: orienting, vigilance, and maintenance versus manipulation-functions of working memory, *Neurosci. Biobehav. Rev.* 26 (2002) 697–712.
- [13] V.L. Brumm, C. Azen, R.A. Moats, A.M. Stern, C. Broomand, M.D. Nelson, R. Koch, Neuropsychological outcome of subjects participating in the PKU adult collaborative study: a preliminary review, *J. Inherit. Metab. Dis.* 27 (2004) 549–566.
- [14] S. Channon, G. Goodman, S. Zlotowitz, C. Mockler, P.J. Lee, Effects of dietary management of phenylketonuria on long-term cognitive outcome, *Arch. Dis. Child.* 92 (2007) 213–218.
- [15] S. Channon, C. Mockler, P. Lee, Executive functioning and speed of processing in phenylketonuria, *Neuropsychology* 19 (2005) 679–686.
- [16] C. Landvogt, E. Mengel, P. Bartenstein, H.G. Buchholz, M. Schreckenberger, T. Siessmeier, A. Scheurich, R. Feldmann, J. Weglage, P. Cumming, F. Zepp, K.

- Ullrich, Reduced cerebral fluoro-L-dopamine uptake in adult patients suffering from phenylketonuria, *J. Cereb. Blood Flow Metab.* 28 (2008) 824–831.
- [17] R. Feldmann, J. Von Berlepsch, S. Kloska, J. Weglage, B. Koletzko, Neuropsychological impairment in adult patients with treated phenylketonuria, *J. Inherit. Metab. Dis.* 31 (2008) 78 (Abstract).
- [18] R. Wappner, S. Cho, R.A. Krommal, V. Schuett, M.R. Seashore, Management of phenylketonuria for optimal outcome: a review of guidelines for phenylketonuria management and a report of surveys of parents, patients, and clinic directors, *Pediatrics* 104 (1999) e68.
- [19] Phenylketonuria (PKU): screening and management, NIH Consensus Statement 17 (2000) 1–33.
- [20] F.J. van Spronsen, P. Burgard, The truth of treating patients with phenylketonuria after childhood: the basis for a new guideline, *J. Inherit. Metab. Dis.* 31 (2008) 673–679.
- [21] S. Schweitzer-Krantz, P. Burgard, Survey of national guidelines for the treatment of phenylketonuria, *Eur. J. Pediatr.* 159 (2000) S70–S73.
- [22] F.J. van Spronsen, K.K. Ahring, M. Gizewska, PKU – what is daily practice in various centres in Europe? Data from a questionnaire by the scientific advisory committee of the European Society of Phenylketonuria and Allied Disorders, *J. Inherit. Metab. Dis.* 32 (2009) 58–64.
- [23] K. Ahring, A. Belanger-Quintana, K. Dokoupil, H. Gokmen Ozel, A.M. Lammardo, A. MacDonald, K. Motzfeldt, M. Nowacka, M. Robert, M. van Rijn, Dietary management practices in phenylketonuria across European centres, *Clin. Nutr.* 28 (2009) 231–236.
- [24] E. Simon, M. Schwarz, J. Roos, N. Dragano, M. Geraedts, J. Siegrist, G. Kamp, U. Wendel, Evaluation of quality of life and description of the sociodemographic state in adolescent and young adult patients with phenylketonuria (PKU), *Health Qual. Life Outcomes* 6 (2008) 25.
- [25] P.B. Acosta, S. Yannicelli, R. Singh, L.J. Eisas 2nd, M.J. Kennedy, L. Bernstein, F. Rohr, C. Trahms, R. Koch, J. Breck, Intake and blood levels of fatty acids in treated patients with phenylketonuria, *J. Pediatr. Gastroenterol. Nutr.* 33 (2001) 253–259.
- [26] P.B. Acosta, S. Yannicelli, R. Singh, S. Mofidi, R. Steiner, E. DeVincentis, E. Jurecki, L. Bernstein, S. Gleason, M. Chetty, B. Rouse, Nutrient intakes and physical growth of children with phenylketonuria undergoing nutrition therapy, *J. Am. Diet. Assoc.* 103 (2003) 1167–1173.
- [27] P.B. Acosta, S. Yannicelli, R.H. Singh, L.J. Eisas 2nd, S. Mofidi, R.D. Steiner, Iron status of children with phenylketonuria undergoing nutrition therapy assessed by transferrin receptors, *Genet. Med.* 6 (2004) 96–101.
- [28] B. Schwahn, E. Mokov, K. Scheidhauer, B. Lettgen, E. Schonau, Decreased trabecular bone mineral density in patients with phenylketonuria measured by peripheral quantitative computed tomography, *Acta Paediatr.* 87 (1998) 61–63.
- [29] D. Modan-Moses, I. Vered, G. Schwartz, Y. Anikster, S. Abraham, R. Segev, O. Efrati, Peak bone mass in patients with phenylketonuria, *J. Inherit. Metab. Dis.* 30 (2007) 202–208.
- [30] D.M. Ney, S.T. Gleason, S.C. van Calcar, E.L. MacLeod, K.L. Nelson, M.R. Etzel, G.M. Rice, J.A. Wolff, Nutritional management of PKU with glycomacropeptide from cheese whey, *J. Inherit. Metab. Dis.* 32 (2009) 32–39.
- [31] J.H. Walter, F.J. White, S.K. Hall, A. MacDonald, G. Rylance, A. Boneh, D.E. Francis, G.J. Shortland, M. Schmidt, A. Vail, How practical are recommendations for dietary control in phenylketonuria?, *Lancet* 360 (2002) 55–57.
- [32] C. Bilginsoy, N. Waitzman, C.O. Leonard, S.T. Ernst, Living with phenylketonuria: perspectives of patients and their families, *J. Inherit. Metab. Dis.* 28 (2005) 639–649.
- [33] K. Peterson, R. Slover, S. Gass, W.K. Seltzer, L.L. McCabe, E.R. McCabe, Blood phenylalanine estimation for the patient with phenylketonuria using a portable device, *Biochem. Med. Metab. Biol.* 39 (1988) 98–104.
- [34] Z. Wang, Y.Z. Chen, S. Zhang, Z. Zhou, Investigation of a phenylalanine-biosensor system for phenylketonuria detection, *Conf. Proc. IEEE Eng. Med. Biol. Soc.* 2 (2005) 1913–1916.
- [35] S. Safos, T.M. Chang, Enzyme replacement therapy in ENU2 phenylketonuric mice using oral microencapsulated phenylalanine ammonia-lyase: a preliminary report, *Artif. Cells Blood Substit. Immobil. Biotechnol.* 23 (1995) 681–692.
- [36] C.N. Sarkissian, Z. Shao, F. Blain, R. Peevers, H. Su, R. Heft, T.M. Chang, C.R. Scriver, A different approach to treatment of phenylketonuria: phenylalanine degradation with recombinant phenylalanine ammonia lyase, *Proc. Natl. Acad. Sci. USA* 96 (1999) 2339–2344.
- [37] J. Liu, X. Jia, J. Zhang, H. Xiang, W. Hu, Y. Zhou, Study on a novel strategy to treatment of phenylketonuria, *Artif. Cells Blood Substit. Immobil. Biotechnol.* 30 (2002) 243–257.
- [38] C.N. Sarkissian, A. Gamez, L. Wang, M. Charbonneau, P. Fitzpatrick, J.F. Lemontt, B. Zhao, M. Vellard, S.M. Bell, C. Henschell, A. Lambert, L. Tsuruda, R.C. Stevens, C.R. Scriver, Preclinical evaluation of multiple species of PEGylated recombinant phenylalanine ammonia lyase for the treatment of phenylketonuria, *Proc. Natl. Acad. Sci. USA* 105 (2008) 20894–20899.
- [39] A. Gamez, L. Wang, C.N. Sarkissian, D. Wendt, P. Fitzpatrick, J.F. Lemontt, C.R. Scriver, R.C. Stevens, Structure-based epitope and PEGylation sites mapping of phenylalanine ammonia-lyase for enzyme substitution treatment of phenylketonuria, *Mol. Genet. Metab.* 91 (2007) 325–334.
- [40] R. Matalon, S. Surendran, K.M. Matalon, S. Tyring, M. Quast, W. Jinga, E. Ezell, S. Szucs, Future role of large neutral amino acids in transport of phenylalanine into the brain, *Pediatrics* 112 (2003) 1570–1574.
- [41] J. Pietz, R. Kreis, A. Rupp, E. Mayatepek, D. Rating, C. Boesch, H.J. Bremer, Large neutral amino acids block phenylalanine transport into brain tissue in patients with phenylketonuria, *J. Clin. Invest.* 103 (1999) 1169–1178.
- [42] J.C. Rocha, F. Martel, Large neutral amino acids supplementation in phenylketonuric patients, *J. Inherit. Metab. Dis.* 32 (2009) 472–480.
- [43] V. Berger, Y. Larondelle, A. Trouet, Y.J. Schneider, Transport mechanisms of the large neutral amino acid L-phenylalanine in the human intestinal epithelial caco-2 cell line, *J. Nutr.* 130 (2000) 2780–2788.
- [44] R. Matalon, K. Michals-Matalon, G. Bhatia, A.B. Burlina, A.P. Burlina, C. Braga, L. Fiori, M. Giovannini, E. Grechanina, P. Novikov, J. Grady, S.K. Tyring, F. Guttler, Double blind placebo control trial of large neutral amino acids in treatment of PKU: effect on blood phenylalanine, *J. Inherit. Metab. Dis.* 30 (2007) 153–158.
- [45] R. Matalon, K. Michals-Matalon, G. Bhatia, E. Grechanina, P. Novikov, J.D. McDonald, J. Grady, S.K. Tyring, F. Guttler, Large neutral amino acids in the treatment of phenylketonuria (PKU), *J. Inherit. Metab. Dis.* 29 (2006) 732–738.
- [46] A. MacDonald, G. Rylance, S.K. Hall, D. Asplin, I.W. Booth, Factors affecting the variation in plasma phenylalanine in patients with phenylketonuria on diet, *Arch. Dis. Child.* 74 (1996) 412–417.
- [47] M.R. Crone, F.J. van Spronsen, K. Oudshoorn, J. Bekhof, G. van Rijn, P.H. Verkerk, Behavioural factors related to metabolic control in patients with phenylketonuria, *J. Inherit. Metab. Dis.* 28 (2005) 627–637.
- [48] P. Vajro, P. Strisciuglio, D. Houssin, G. Huault, J. Laurent, F. Alvarez, O. Bernard, Correction of phenylketonuria after liver transplantation in a child with cirrhosis, *N. Engl. J. Med.* 329 (1993) 363.
- [49] M.E. Blaskovics, G.E. Schaeffler, S. Hack, Phenylalaninaemia. Differential diagnosis, *Arch. Dis. Child.* 49 (1974) 835–843.
- [50] N. Longo, Disorders of bipterin metabolism, *J. Inherit. Metab. Dis.* 32 (2009) 333–342.
- [51] A. Ponzone, O. Guardamagna, M. Spada, S. Ferraris, R. Ponzone, L. Kierat, N. Blau, Differential diagnosis of hyperphenylalaninaemia by a combined phenylalanine-tetrahydrobiopterin loading test, *Eur. J. Pediatr.* 152 (1993) 655–661.
- [52] N. Blau, B. Thöny, M. Spada, A. Ponzone, Tetrahydrobiopterin and inherited hyperphenylalaninemia, *Turk. J. Pediatr.* 38 (1996) 19–35.
- [53] S. Kure, D.C. Hou, T. Ohura, H. Iwamoto, S. Suzuki, N. Sugiyama, O. Sakamoto, K. Fujii, Y. Matsubara, K. Narisawa, Tetrahydrobiopterin-responsive phenylalanine hydroxylase deficiency, *J. Pediatr.* 135 (1999) 375–378.
- [54] A.C. Muntau, W. Roschinger, M. Habich, H. Demmelmair, B. Hoffmann, C.P. Sommerhoff, A.A. Roscher, Tetrahydrobiopterin as an alternative treatment for mild phenylketonuria, *N. Engl. J. Med.* 347 (2002) 2122–2132.
- [55] N. Blau, H. Erlandsen, The metabolic and molecular bases of tetrahydrobiopterin-responsive phenylalanine hydroxylase deficiency, *Mol. Genet. Metab.* 82 (2004) 101–111.
- [56] H. Levy, B. Burton, S. Cederbaum, C. Scriver, Recommendations for evaluation of responsiveness to tetrahydrobiopterin (BH(4)) in phenylketonuria and its use in treatment, *Mol. Genet. Metab.* 92 (2007) 287–291.
- [57] N. Blau, A. Belanger-Quintana, M. Demirkol, F. Feillet, M. Giovannini, A. MacDonald, F.K. Trefz, F.J. van Spronsen, Optimizing the use of sapropterin (BH(4)) in the management of phenylketonuria, *Mol. Genet. Metab.* 96 (2009) 158–163.
- [58] B.K. Burton, D.K. Grange, A. Milanowski, G. Vockley, F. Feillet, E.A. Crombez, V. Abadie, C.O. Harding, S. Cederbaum, D. Dobbelaere, A. Smith, A. Dorembaum, The response of patients with phenylketonuria and elevated serum phenylalanine to treatment with oral sapropterin dihydrochloride (6R-tetrahydrobiopterin): a phase II, multicentre, open-label, screening study, *J. Inherit. Metab. Dis.* 30 (2007) 700–707.
- [59] H.L. Levy, A. Milanowski, A. Chakrapani, M. Cleary, P. Lee, F.K. Trefz, C.B. Whitley, F. Feillet, A.S. Feigenbaum, J.D. Bechuk, H. Christ-Schmidt, A. Dorenbaum, Sapropterin Research Group, Efficacy of sapropterin dihydrochloride (tetrahydrobiopterin, 6R-BH₄) for reduction of phenylalanine concentration in patients with phenylketonuria: a phase III randomised placebo-controlled study, *Lancet* 370 (2007) 504–510.
- [60] S.W. Gersting, K.F. Kemter, M. Staudigl, D.D. Messing, M.K. Danecka, F.B. Lagler, C.P. Sommerhoff, A.A. Roscher, A.C. Muntau, Loss of function in phenylketonuria is caused by impaired molecular motions and conformational instability, *Am. J. Hum. Genet.* 83 (2008) 5–17.
- [61] S.C. Kwok, F.D. Ledley, A.G. DiLella, K.J. Robson, S.L. Woo, Nucleotide sequence of a full-length complementary DNA clone and amino acid sequence of human phenylalanine hydroxylase, *Biochemistry* 24 (1985) 556–561.
- [62] F.D. Ledley, H.E. Grenett, A.G. DiLella, S.C. Kwok, S.L. Woo, Gene transfer and expression of human phenylalanine hydroxylase, *Science* 228 (1985) 77–79.
- [63] R.C. Eisensmith, S.L. Woo, Gene therapy for phenylketonuria, *Eur. J. Pediatr.* 155 (1996) S16–S19.
- [64] S.E. Raper, N. Chirmule, F.S. Lee, N.A. Wivel, A. Bagg, G.P. Gao, J.M. Wilson, M.L. Batshaw, Fatal systemic inflammatory response syndrome in an ornithine transcarbamylase deficient patient following adenoviral gene transfer, *Mol. Genet. Metab.* 80 (2003) 148–158.
- [65] C.O. Harding, M.B. Gillingham, K. Hamman, H. Clark, E. Goebel-Daghighi, A. Bird, D.D. Koeberl, Complete correction of hyperphenylalaninemia following liver-directed, recombinant AAV2/8 vector-mediated gene therapy in murine phenylketonuria, *Gene Ther.* 13 (2006) 457–462.
- [66] C. Harding, Progress toward cell-directed therapy for phenylketonuria, *Clin. Genet.* 74 (2008) 97–104.
- [67] I.E. Alexander, S.C. Cunningham, G.J. Logan, J. Christodoulou, Potential of AAV vectors in the treatment of metabolic disease, *Gene Ther.* 15 (2008) 831–839.

- [68] G.M. Enns, M.T. Millan, Cell-based therapies for metabolic liver disease, *Mol. Genet. Metab.* 95 (2008) 3–10.
- [69] Z. Ding, C.O. Harding, A. Rebuffat, L. Elzaouk, J.A. Wolff, B. Thöny, Correction of murine PKU following AAV-mediated intramuscular expression of a complete phenylalanine hydroxylating system, *Mol. Ther.* 16 (2008) 673–681.
- [70] C.R. Scriver, The PAH gene, phenylketonuria, and a paradigm shift, *Hum. Mutat.* 28 (2007) 831–845.
- [71] F.J. van Spronsen, M. Hoeksma, D.J. Reijngoud, Brain dysfunction in phenylketonuria: is phenylalanine toxicity the only possible cause?, *J. Inherit. Metab. Dis.* 32 (2009) 46–51.
- [72] L.B. Möller, M. Paulsen, R. Koch, R. Moats, P. Guldborg, F. Guttler, Inter-individual variation in brain phenylalanine concentration in patients with PKU is not caused by genetic variation in the 4F2hc/LAT1 complex, *Mol. Genet. Metab.* 86 (2005) S119–S123.
- [73] J. Weglage, D. Wiedermann, J. Denecke, R. Feldmann, H.G. Koch, K. Ullrich, E. Harms, H.E. Moller, Individual blood–brain barrier phenylalanine transport determines clinical outcome in phenylketonuria, *Ann. Neurol.* 50 (2001) 463–467.
- [74] R. Koch, R. Moats, F. Guttler, P. Guldborg, M. Nelson Jr., Blood–brain phenylalanine relationships in persons with phenylketonuria, *Pediatrics* 106 (2000) 1093–1096.
- [75] R.A. Moats, R. Koch, K. Moseley, P. Guldborg, F. Guttler, R.G. Boles, M.D. Nelson Jr., Brain phenylalanine concentration in the management of adults with phenylketonuria, *J. Inherit. Metab. Dis.* 23 (2000) 7–14.
- [76] R. Koch, K.D. Moseley, S. Yano, M. Nelson Jr., R.A. Moats, Large neutral amino acid therapy and phenylketonuria: a promising approach to treatment, *Mol. Genet. Metab.* 79 (2003) 110–113.
- [77] H.N. Christensen, Metabolism of amino acids and proteins, *Annu. Rev. Biochem.* 22 (1953) 233–260.
- [78] A.E. Andersen, L. Avins, Lowering brain phenylalanine levels by giving other large neutral amino acids. A new experimental therapeutic approach to phenylketonuria, *Arch. Neurol.* 33 (1976) 684–686.
- [79] O.E. Pratt, A new approach to the treatment of phenylketonuria, *J. Ment. Defic. Res.* 24 (1980) 203–217.
- [80] C.A. Brass, O. Greengard, Modulation of cerebral catecholamine concentrations during hyperphenylalaninaemia, *Biochem. J.* 208 (1982) 765–771.
- [81] M.K. Jordan, R.L. Brunner, M.M. Hunt, H.K. Berry, Preliminary support for the oral administration of valine, isoleucine, and leucine for phenylketonuria, *Dev. Med. Child Neurol.* 27 (1985) 33–39.
- [82] S. Schindeler, S. Ghosh-Jerath, S. Thompson, A. Rocca, P. Joy, A. Kemp, C. Rae, K. Green, B. Wilcken, J. Christodoulou, The effects of large neutral amino acid supplements in PKU: an MRS and neuropsychological study, *Mol. Genet. Metab.* 91 (2007) 48–54.
- [83] G. Kapatos, S. Kaufman, Peripherally administered reduced pterins do enter the brain, *Science* 212 (1981) 955–956.
- [84] L. Miller, T. Insel, M. Scheinin, J. Aloï, D.L. Murphy, M. Linnoila, W. Lovenberg, Tetrahydrobiopterin administration to rhesus macaques. Its appearance in CSF and effect on neurotransmitter synthesis, *Neurochem. Res.* 11 (1986) 291–298.
- [85] S. Kaufman, G. Kapatos, R.R. McInnes, J.D. Schulman, W.B. Rizzo, Use of tetrahydropterins in the treatment of hyperphenylalaninemia due to defective synthesis of tetrahydrobiopterin: evidence that peripherally administered tetrahydropterins enter the brain, *Pediatrics* 70 (1982) 376–380.
- [86] B. Thöny, A.C. Calvo, T. Scherer, R.M. Svebak, J. Haavik, N. Blau, A. Martinez, Tetrahydrobiopterin shows chaperone activity for tyrosine hydroxylase, *J. Neurochem.* 106 (2008) 672–681.